

Influence of MRI examinations on animal welfare and study results

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Short title: MRI effect on mouse welfare and study results

Influence of MRI examinations on animal welfare and study results

Abstract

Objectives: MRI is considered to be well tolerated by laboratory animals. However, no systematic study has been performed yet, proving this assumption. Therefore, the aim of this study was to investigate possible effects of longitudinal native and contrast-enhanced (CE) 1T and 7T MRI examinations on mouse welfare as well as 4T1 breast cancers progression and therapy response.

Material and Methods: 47 healthy and 72 breast cancer-bearing mice (4T1) were investigated. 1T (ICON) and 7T (Biospec) MRI measurements were performed thrice per week under isoflurane anesthesia in healthy BALB/c mice for four weeks and three times within two weeks in tumor-bearing animals. Animal welfare was examined by an observational score sheet, rotarod performance, heart rate measurements and assessment of fecal corticosterone metabolites (FCMs). Furthermore, we investigated, whether CE MRI influences the study outcome. Therefore, hemograms and organ weights were obtained and 4T1 tumor-growth, perfusion, immune cell infiltration, as well as response to the multikinase inhibitor regorafenib were investigated. Statistical comparisons between groups were performed using ANOVA and Tukey's or Bonferroni post-hoc tests.

Results: Mice showed no alterations in the observational score sheet rating, rotarod performance, heart rate and FCMs ($p > 0.05$) after repeated MRI at both field strengths. However, spleen weights were reduced in all healthy mouse groups that received isoflurane anesthesia ($p < 0.001$) including the groups investigated by 1T and 7T MRI ($p = 0.02$). Neither tumor progression nor response to the regorafenib treatment were affected by isoflurane anesthesia or CE MRI monitoring. Furthermore, immunohistological tumor analysis did not indicate an effect of isoflurane and MRI on macrophage infiltration of tumors, perfusion of tumor vessels, and apoptotic cell rate ($p > 0.05$).

Conclusion: Repeated MR imaging did not influence the welfare of mice and did not affect tumor growth and therapy response of 4T1 tumors. However, systemic immunological effects of isoflurane anesthesia need to be considered to prevent potential bias.

Keywords: MRI, animal welfare, severity assessment, long-term effects, regorafenib, isoflurane, anesthesia, behavior, rotarod, breast cancer

Abbreviations:

BL = Baseline, FCMs = Fecal Corticosterone Metabolites, EPI = Echo-Planar-Imaging, CE = Contrast enhanced

Introduction

Non-invasive imaging is broadly used in (pre-)clinical studies to depict morphological and (patho-) physiological processes^{1,2} and is often argued to contribute to the 3Rs³. These were initially described in 1959 by Russel and Burch and define the three main principles for the ethically correct handling of laboratory animals. In detail, they stand for Replacement, Reduction and Refinement. This can be achieved by reducing the number of animals needed, refining the studies in favor of animal welfare and data quality and by replacing animal experiments, e.g. by in vitro techniques. Non-invasive imaging contributes to the 3Rs by enabling longitudinal studies of the same animal, thus reducing the number of animals and allowing intraindividual analyses⁴. However, longitudinal examinations are also discussed to induce higher burden due to suffering of the single animal for a longer period. Although imaging techniques are well tolerated in humans, in laboratory animals few important differences need to be highlighted. First, animal handling itself can cause anxiety⁵ and alter physiological parameters⁶. Second, animals need to be anesthetized throughout the whole imaging procedure. In this context, isoflurane is one of the most commonly used anesthetic agents. It acts on the γ -aminobutyric acid and glutamatergic receptors in the brain, but the mechanism of action is not yet fully known⁷. With respect to animal behavior, it was shown to decrease burrowing and open field exploratory behavior in C57BL/6JRj mice⁸, and significantly modulated immune responses⁹. Furthermore, isoflurane was shown to alter plasma corticosterone levels in the hippocampus¹⁰ as well as activate TrkB and can therefore have an antidepressant effect¹¹. These risks increase with anesthesia length and repetitions⁸ and are consequently especially relevant for MRI¹².

Next to effects related to the anesthesia, preclinical MRI is typically performed at higher field strengths to achieve sufficient spatial resolution¹³ as well as more often and in shorter time intervals¹² compared to clinical investigations. High noise levels¹⁴ and the magnetic field itself can influence animals' behavior and even cause depressive symptoms¹⁵. Especially ultrafast echo-

planar-imaging (EPI) sequences require high gradient amplitudes and fast switching rates, which can evoke peripheral nerve stimulation^{16,17} or tissue heating¹⁸ resulting in muscle twitching and even pain¹⁹. Thus, thresholds were defined to prevent sequence-related injuries for humans²⁰. However, in small animal imaging gradient amplitudes above those thresholds are often needed for sufficient spatial and temporal resolution. Depending on the scientific question, MRI can also require the injection of a contrast agent^{21,22}, causing additional stress. Taken together, all aforementioned issues might influence animal welfare and maybe even study results, leading to an interpretation bias.

So far, no systematic study has been published examining the long-term effects of standard MRI protocols on animal welfare and scientific outcome in cancer research. Thus, we investigated the influence of native and contrast-enhanced (CE) MRI under isoflurane anesthesia with different field strengths on welfare and physiological parameters of healthy mice. Furthermore, effects of CE MRI on the progression of highly vascularized 4T1 breast cancers and its response to the multikinase receptor inhibitor regorafenib were examined. This study should demonstrate possible effects of MRI on animal welfare and scientific outcome and moreover reevaluate the suitability of MRI in preclinical research.

Material and Methods

Animal experiments

All animal experiments were approved by the German State Office for Nature, Environment and Consumer Protection (LANUV) North Rhine-Westphalia. Overall, 119 female BALB/cAnNRj mice (age 10-12 weeks; Janvier Labs, Saint Berthevin, France) were housed in groups of 5 animals under specific pathogen free conditions with a 12 h light/dark cycle. The environment was temperature (20-24 °C) and humidity (45-65 %) controlled according to the guidelines of the “Federation for Laboratory Animal Science Associations” (FELASA, www.felasa.eu). Acidified water and standard pellets for laboratory mice (Sniff GmbH, Soest, Germany) were offered ad libitum.

Orthotopic tumor inoculation and anti-tumor therapy

Murine triple-negative breast cancer cells (4T1²³, ATCC® CRL-2539™, Manassas, Virginia, USA; 4x10⁴ cells in 50µl RPMI 1640 cell culture medium) were injected orthotopically into the right mammary fat pad of n = 72 mice. Tumor sizes were assessed daily using caliper measurements and the volume was calculated using the formula “length*width*width*(π/6)”. Tumors with sizes of less than 5 mm³ on day 6 after tumor cell injection were excluded from analyses.

Six days after tumor cell injection, animals were randomly allocated to receive a daily oral dose of either 10 mg/kg body weight regorafenib (multikinase inhibitor, Merck, Darmstadt, Germany) dissolved in polyethyleneglycol 400, 1,2-propandiol and pluronic F68 (all from Merck, Darmstadt, Germany), as described²⁴ or the corresponding amount of vehicle solution.

MRI protocol

Imaging was performed using a 1T (ICON, Bruker, Ettlingen, Germany) or a 7T MRI scanner (BioSpec, Bruker Ettlingen, Germany) with a transceiver mouse volume coil (sequence details in Supplemental Digital Content 1 tables S1 and S2).

Healthy and tumor-bearing mice of the 7T MRI and 7T CE MRI groups were imaged according to a representative MRI protocol performed in our institute: (i) localization of liver and mammary fat pad by transversal T1-RARE sequences, (ii) imaging of both using a T2-RARE sequence, (iii) T1-weighted saturation recovery gradient echo sequence for dynamic CE MRI with 80 measurements and a temporal resolution of 8.4 images per second resulting in a total scan time of 11.2 min. After a baseline measurement of 1 min, all healthy and tumor-bearing animals of the 7T CE MRI group received an intravenous injection of 0.1 mmol/kg body weight Gadovist (Bayer, Germany) between the 7th and 10th repetition of the T1-FLASH sequence for dynamic contrast enhanced imaging.

An additional group of 3 healthy mice underwent 7T MRI of the brain and lung with different EPI sequences varying in TR (667 ms, 990 ms, 3000 ms; see table S2) to investigate acute effects of these sequences on the respiration rate.

Influence of repeated MRI with standard sequences on animal welfare and health

Influence of repeated MRI on animal welfare and health was assessed in n = 44 healthy female BALB/cAnNRj mice. At least 5 animals were randomized to either experimental group: (i) no imaging (control), (ii) isoflurane anesthesia, (iii) 1T MRI, (iv) 7T MRI, and (v) 7T CE MRI. Isoflurane anesthesia alone (30 min, 2 % isoflurane in O₂) or in combination with MRI was carried out 3 times per week for 4 weeks, starting at day 0.

The health state of all animals was monitored daily using a previously described modified score sheet²⁵ (see Supplemental Digital Content 1, table S3). Briefly, alterations in body weight, general state, spontaneous behavior and treatment specific parameters (tumor growth, anti-tumor therapy) were documented and allocated to a point grading system, where 0 points describe no alteration

of the physiological state and ≥ 20 points highest severity and humane endpoint. Before the first imaging session, all animals were trained for behavioral examinations. Baseline values for rotarod performance, heart rate and fecal corticosterone metabolites (FCMs) concentration were assessed. Changes of these parameters were then monitored twice per week (figure 1). On the last day of the experiment, a final assessment of all behavioral tests was performed, and blood was collected before euthanasia.

Rotarod test

The rotarod (Panlab Harvard Apparatus, Barcelona, Spain) test was applied to discover changes in motor coordination and balance. Mice were put on a spinning cylinder, with a starting speed of 4 rotations per minute (rpm), accelerating steadily to a maximum speed of 40 rpm after 5 min. The speed at which the mouse fell of the cylinder was recorded. Each measurement was repeated twice. Baseline values were set to 100 % and percent change of the subsequent time points were calculated on an individual basis.

Assessment of heart rate

Heart rate was measured in conscious mice as an indicator of stress and discomfort (CODA System, Kent Scientific Corporation, Torrington, CT, USA). Therefore, mice were restrained in a plexiglas tube on a tempered panel. The tail was placed inside an occlusion and volume pressure cuff. Heart rate was assessed by volume pressure recording and analyzed using the suppliers' software. Each measurement consisted of 15 repetitions to compensate for movement artifacts. Baseline values were set to 100 %, and percent changes at the subsequent time points were calculated on an individual basis.

FCMs measurements

Feces was collected during every animal handling and stored at -80 °C. Samples of 3 consecutive days were pooled and dried for 24 h at 50 °C. Then, an aliquot of 50mg was dissolved in 80% Methanol (1ml, Merck, Darmstadt, Germany) over night at 4 °C. Samples were homogenized and centrifuged for 10 min at 3000 xg (relative centrifugal acceleration) (Fresco 21 and Pico 21 Heraeus, Hanau, Germany). Subsequently, the samples were analyzed by 5 α -pregnane-3 β ,11 β ,21-triol-20-one enzyme immune assay²⁶ at the Institute of Physiology, Pathophysiology and Experimental Endocrinology of the University of Veterinary medicine in Vienna, Austria. Baseline values were set to 100 % and percent changes at the subsequent time points were calculated on an individual basis.

Influence of Echo-Planar-Imaging on animal welfare and respiratory rate

For the evaluation of acute irritating effects on the central nervous system deriving from EPI sequences, 3 animals were investigated 3 times within one week and respiratory rates were documented every 5s during the scans.

Influence of repeated MRI on tumor-pathophysiology

To test whether repeated MRI influences 4T1 tumor-pathophysiology, n = 72 regorafenib or vehicle treated animals were randomized to the following experimental groups: (i) no imaging, (ii) isoflurane anesthesia, and (iii) CE 7T MRI. Isoflurane anesthesia alone (30 min, 2 % isoflurane in O₂) or in combination with CE MRI was performed on days 7, 10 and 14 after tumor inoculation. Before the first imaging session, all animals were trained, and baseline values assessed as described for healthy animals. Changes were monitored on days 11, 13, and 15 after tumor cell injection (figure 2). On the last day of the experiment, a final assessment of all behavioral tests was performed, and blood was collected before euthanasia.

Hemograms

Blood was collected by retro-bulbar sinus puncture and analyzed for numbers of leukocytes, erythrocytes, thrombocytes, hemoglobin amount and hematocrit concentration with Celltac alpha MEK-6550 (Nihon Kohden, Shinjuku, Japan).

Organ and tumor tissue preparation

Before euthanasia, mice were anesthetized with ketamine/xylazine in 0.9 % NaCl (120 mg/kg body weight ketamine /16 mg/kg body weight xylazine; 30 µl/10 g body weight intraperitoneal injection). All tumor-bearing mice were injected intravenously with rhodamine labeled *Ricinus Communis* agglutinin I (15 mg/kg body weight; Vector Laboratories, Burlingame, CA, USA) to stain perfused tumor vessels. After 15 min mice hearts were perfused with 10 ml phosphate buffer saline (Merck, Darmstadt, Germany) through the left ventricle of the heart. Organs of interest (brain, heart, lungs, liver, spleen, kidneys) were macroscopically examined for abnormalities, and weighted. Tumors were excised, weighted, embedded in Tissue Tek[®] O.C.T. Compound (Sakura, Alphen aan den Rijn, Netherlands) and stored at -80 °C.

Immunohistology

Tumors were cryosectioned into 8 µm thick slices (CM3050S, Leica, Wetzlar, Germany). For immunohistological stainings tumor slices were fixed with methanol (80 %, Merck, Darmstadt, Germany) and ice-cold acetone (100 %, Merck, Darmstadt, Germany). Blood vessels were stained with rat-anti-mouse PECAM-1 monoclonal CD31 (50 µg/ml, BD Bioscience, San Jose, CA, USA, #553370), followed by donkey-anti-rat IgG (H+L) (0.001 µg/µl, #712-546-153 Dianova, Hamburg, Germany). Macrophages were stained with F4/80 rat-anti-mouse antibody (20 µg/ml, #MCA497GA Bio-Rad, Hercules, CA, USA), followed by donkey-anti-rat IgG (H+L) (0,001 µg/µl, #712-546-153 Dianova, Hamburg, Germany). For detection of apoptotic cells, tumor slices were fixed with 4 % paraformaldehyde and stained using the in-situ cell death detection kit labelled with fluorescein

(TUNEL, Roche, Basel, Switzerland). All slices were counterstained with 4',6-diamidino-2-phenylindole (0.5 µg/ml DAPI, Merck, Darmstadt, Germany) to visualize nuclei. Five fluorescent micrographs per section were captured using the Axio Imager.M2 microscope (Zeiss, Göttingen, Germany) with a high-resolution camera (AxioCam MRm Rev.3, Zeiss, Göttingen, Germany) and quantified using ImageJ2²⁷ (National Institute of Health, Bethesda, MD, USA).

Vessel perfusion was quantified by determining the percentage of co-localized CD31 and *Ricinus Communis* agglutinin I positive area fraction. The positive area fraction of F4/80⁺ cells and TUNEL⁺ cells was determined and the relation to DAPI⁺ cells was calculated to quantify the percentage of macrophages and apoptotic cells, respectively.

Statistical analysis

Statistical analysis was performed with SPSS (IBM Corp., v25, Sanborn, NY, USA, academic license) and GraphPad Prism5 (Graphpad Software, v5.01, San Diego, CA, USA, academic license). Data were tested for normality and analyzed with One-Way ANOVA and Tukey`s post-hoc test or repeated measures ANOVA with Bonferroni post-hoc test on a 95 % confidence interval. p-values < 0.05 were considered statistically significant. All data are presented as mean ± standard deviation.

Results

Influence of repeated MRI on animal welfare and health

First, the influence of repeated MRI at different field strengths with and without contrast agent was studied in healthy BALB/c mice. The general burden of each animal was rated daily by evaluation of several welfare parameters defined in a score sheet (Supplemental Digital Content 1 table S3). In all groups, scores did not exceed a mild burden throughout the experiments (figure 3A).

In line with this, no alteration of the rotarod performance could be detected after repeated anesthesia or MRI when compared to baseline values ($p = 0.086$; table 1; figure 3B). Furthermore, stress associated parameters like heart rate ($p = 0.272$; table 1; figure 3C) and FCMs ($p = 0.117$; table 1; figure 3D) did not show any alteration after repeated anesthesia or MRI.

Hemograms, assessed on the last day of the experiment, also revealed no deterioration of the animals' health reflected by a comparable number of leukocytes (figure 3E), erythrocytes, thrombocytes, and hemoglobin as well as hematocrit values in the different experimental groups (Supplemental Digital Content 1, table S4). However, it needs to be mentioned that 5 animals per group of the control, isoflurane, and 7T MRI group had to be excluded from analysis due to an internal error of the measurement device. Statistical analysis was performed with the remaining 5 mice for the control, isoflurane, and 7T group.

Finally, gross necropsy of brain, lungs, heart, liver, spleen and kidneys did not show any macroscopic abnormalities. However, the spleen weights of control mice were significantly higher ($p < 0.001$) compared to all other experimental groups of healthy mice that received isoflurane anesthesia alone or in combination with MRI (control: 0.109 ± 0.007 g, isoflurane: 0.096 ± 0.007 g, 1T MRI: 0.096 ± 0.009 g, 7T MRI: 0.091 ± 0.009 g, CE 7T MRI: 0.095 ± 0.01 g; figure 3F). Importantly, no significant difference was observed in case of the net isoflurane group compared to the imaging groups (that are anesthetized during the measurements with isoflurane), indicating that the decrease in spleen weights is solely an effect of inhalation anesthesia.

Influence of EPI sequences on animal welfare and respiratory rate

Within a pilot study, the influence of EPI MRI on the respiratory rate and animal welfare was investigated in 3 animals. In line with our results of repeated MRI, EPI sequences had no impact on rotarod performance and heart rate measurements in healthy mice. Furthermore, no changes of respiratory rates were assessed during the measurements that would indicate an irritation of the brainstem (Supplemental Digital Content 1, figure S1). Since no alterations were observed, no additional animals were examined for ethical reasons.

Influence of repeated MRI on tumor progression and therapy response

Next, the influence of repeated CE MRI on progression and therapy response of 4T1 murine breast cancers was investigated. Additionally, the animals' wellbeing was assessed as described for healthy mice.

The tumors of regorafenib treated animals were significantly smaller compared to vehicle treated tumors from day 11 after tumor cell injection on ($p = 0.015$) (figure 4A-B). The therapeutic effect of regorafenib on 4T1 tumors could be confirmed by histological analysis showing a slightly ($p > 0.05$) lower macrophage infiltration (figure 4D) and a considerably higher percentage of apoptotic cells in regorafenib compared to vehicle treated tumors (figure 4E). However, no alteration in vessel perfusion (figure 4C) was found. Furthermore, no influence of isoflurane anesthesia or MRI on tumor growth could be detected in neither vehicle ($p = 0.291$) nor regorafenib treated ($p = 0.930$) animals. In this line, isoflurane and MRI did not change perfusion, macrophage infiltration and apoptosis in tumors treated with vehicle or regorafenib (figure 4C-E).

Score sheet evaluation of tumor-bearing animals revealed higher scores as compared to healthy animals. Nevertheless, the burden exceeded the mild level only for a few individual animals on the last day of the experiment, which could be attributed to cancer progression (Supplemental Digital Content 1 figure S2A). Treatment of tumor-bearing mice with regorafenib had no influence on animal welfare assessed by score sheet evaluation when compared to vehicle treated ones.

Furthermore, neither isoflurane anesthesia nor MRI altered welfare of tumor-bearing animals according to the point grading system from our score sheet evaluation (table 2). In this line, rotarod performance ($p = 0.093$; table 2; figure S2B), heart rate ($p = 0.924$; table 2; figure S2C) and FCMs ($p = 0.089$, table 2, figure S2D) did not differ in tumor-bearing mice indicating that tumor growth, regorafenib therapy and isoflurane anesthesia as well as MRI did not affect motor coordination and stress levels, and therefore, animal welfare (figure S2B-D).

In tumor-bearing mice, the hemograms showed no differences in the number of erythrocytes, hemoglobin or hematocrit content compared to healthy mice. However, leukocyte counts were significantly higher ($p = 0.002$) and the number of thrombocytes significantly reduced ($p < 0.001$) (table S5). Within tumor-bearing groups, regorafenib therapy significantly reduced the number of leukocytes ($p = 0.019$) (figure 5A). In vehicle treated animals, isoflurane anesthesia as well as MRI resulted in a considerable reduction ($p = 0.093$) of leukocyte numbers, whereas leukocyte counts in regorafenib treated animals remained unchanged ($p = 0.239$) (vehicle: control: $69.45 \pm 38.58 \times 10^3/\mu\text{l}$, isoflurane: $53.07 \pm 27.38 \times 10^3/\mu\text{l}$, CE MRI: $38.57 \pm 25.09 \times 10^3/\mu\text{l}$; regorafenib: control: $43.29 \pm 26.5 \times 10^3/\mu\text{l}$, isoflurane: $40.88 \pm 12.59 \times 10^3/\mu\text{l}$, CE MRI: $32.32 \pm 16.40 \times 10^3/\mu\text{l}$) (figure 5A).

Furthermore, spleen weights were significantly higher in tumor bearing mice in comparison to healthy animals ($p < 0.001$) and regorafenib resulted in a significant reduction in spleen weights compared to vehicle controls (figure 5B) ($p = 0.007$). In addition, the spleen weights of the vehicle-treated cohort were reduced in the isoflurane group and significantly reduced in the CE MRI group ($p = 0.022$) (vehicle: control: 0.298 ± 0.071 g, isoflurane: 0.252 ± 0.05 g, CE MRI: 0.219 ± 0.077 g; regorafenib: control: 0.220 ± 0.063 g, isoflurane: 0.236 ± 0.053 g, CE MRI: 0.186 ± 0.031 g) (figure 5B).

Discussion

There is a general agreement of researchers and regulatory boards that the number of laboratory animals and their suffering should be minimized. Although, imaging procedures are assigned to the mild severity category²⁸, hardly any data are available about imaging related suffering or effects on study results. Our results of score sheet evaluations in healthy animals indicate that even after longitudinal MRI animal burden does not exceed a mild range. Furthermore, rotarod performance, heart rate and FCMs analysis indicated no long-term effects induced by MRI. In line with our findings, the few preliminary studies showed no effects of magnetic fields²⁹⁻³¹ or MRI³² between 0.15T and 6.3T on exploratory behavior, locomotion, memory or fetal development in rats and mice, when exposed 20-120 min to a magnetic field. However, also contrary results can be found describing e.g. circling behavior after 30 min exposure of rats to 7 or 14T fields¹⁵. Comparable findings were reported for mice along with conditioned taste aversions³³. However, in the aforementioned studies, animals were awake and effects on their behavior were investigated directly after magnetic field exposure. In the present study, our aim was to explore the effects of a standard MRI setup with T_{1w} and T_{2w} sequences, which includes isoflurane anesthesia. Thus, these acute effects could not be captured due to the well-known anesthesia induced behavioral alterations in the immediate post-anesthetic phase⁸. However, it is noteworthy that also in the group exposed to EPI MRI sequences, no changes in the respiratory rate, pointing to irritation of the brainstem, could be seen. The latter was also assumed to be responsible for the circling behavior and taste aversion in rats found by Houpt and coworkers¹⁵.

Despite these encouraging results for the use of MRI, we need to mention, as a limitation of our study, that the rotarod test only measures motor coordination and balance and might not be most sensitive for measuring anxiety or stress³⁴. Thus, future studies need to comprise further tests like the open field, combining visualization of motor function with a sensitive analysis of exploratory behavior of individual animals³⁵.

With respect to physiological alterations, healthy mice showed a decrease in spleen weights after isoflurane anesthesia independent of MRI. Comparable findings were reported for acepromazine and propofol administration in dogs³⁶, but were not yet described after repeated isoflurane anesthesia in mice. However, isoflurane has an impact on the immune system and is known to decrease the numbers of B- and T-cells (both present in spleen) in humans⁹ and mice³⁷.

In line with our results on healthy animals, isoflurane anesthesia and MRI had no impact on animal welfare in 4T1 tumor-bearing mice. The score sheet evaluation showed an elevated total score at the end of the experiment up to a moderate range caused by cancer progression. In this context, tumor growth induced an immune response resulting in increased leukocyte counts and spleen weights compared to healthy mice, which is in line with the literature³⁸. Treatment of tumor-bearing mice with regorafenib slightly reduced leukocyte counts and spleen weights most likely due to myelosuppression³⁹. As in healthy mice, lower spleen weights were measured in vehicle treated animals after isoflurane anesthesia, whereas this effect could not be observed in regorafenib treated mice. In the latter group, the reduction in spleen weights induced by regorafenib may have masked the isoflurane related effect.

With respect to disease progression, regorafenib significantly inhibited 4T1 tumor-growth, however, the imaging procedure itself (with contrast agent) had no detectable effect on tumor physiology nor growth nor therapy response. Even though there are no reports on magnetic fields influencing tumor growth in mice, this needs to be considered since a reduction of tumor size in male hamsters has been described after exposure to a 586 mT static magnetic field for 3 h⁴⁰. Additionally, several effects of intravenous (e.g. ketamine) and volatile (e.g. isoflurane) anesthetics on tumor development and the immune system were described caused by e.g. enhancing hypoxia inducible factor-1 α activity, apoptotic resistance of tumor cells or attenuation of natural killer cell activity⁴¹. In this context, it is a limitation of our study, that only female mice were included, since tumor response to isoflurane might be sex-dependent. For example, administration of isoflurane resulted in faster melanoma growth in male, but not in female mice, possibly by sex-dependent differences in their

immune response⁴². However, since our study focused on orthotopic breast cancers the choice of female mice was reasonable.

In conclusion, we systematically evaluated potential risks and limitations of repeated MRI with regard to animal welfare and cancer research. Based on our findings we consider MRI a safe tool for longitudinal functional and morphological investigations with respect to animal welfare. By doing this, we hopefully erase concerns associated with preclinical repeated MRI of small laboratory animals. However, the influence of isoflurane on the immune system has to be considered especially in immunotherapy related research.

References

1. Fischman AJ, Alpert NM, Rubin RH. Pharmacokinetic imaging: a noninvasive method for determining drug distribution and action. *Clin Pharmacokinet* 2002;41(8):581-602.
2. Taoka T, Jost G, Frenzel T, et al. Impact of the Glymphatic System on the Kinetic and Distribution of Gadolinium in the Rat Brain. *Invest Radiol* 2018;53(9):529-534.
3. Russel WMS, Burch RL. The principles of humane experimental technique. Wheathampstead (UK) 1959: Universities Federation for Animal Welfare.
4. Beckmann N, Ledermann B. Noninvasive small rodent imaging: Significance for the 3R principles. In: Kiessling F, Pichler B, Hauff P (eds) *Small Animal Imaging*. Cham, 2017; 69-87.
5. Hurst JL, West RS. Taming anxiety in laboratory mice. *Nat Methods* 2010;7:825-826.
6. Swaim LD, Taylor HW, Jersey GC. The effect of handling techniques on serum alt activity in mice. *J Appl Toxicol* 1985;5(3):160-162.
7. Ranft A, Kurz J, Deuringer M, et al. Isoflurane modulates glutamatergic and GABAergic neurotransmission in the amygdala. *Eur J Neurosci* 2004;20:1276-1280.
8. Hohlbaum K, Bert B, Dietze S, et al. Severity classification of repeated isoflurane anesthesia in C57BL/6JRj mice – Assessing the degree of distress. *PLoS ONE* 2017;12(6).

9. Stollings LM, Jia LJ, Tang P, et al. Immune modulation by volatile anesthetics. *Anesthesiology* 2016;125(2):399-411.
10. Bekhbat M, Merrill L, Kelly SD, et al. Brief anesthesia by isoflurane alters plasma corticosterone levels distinctly in male and female rats: Implication for tissue collection methods. *Behav Brain Res* 2016;305:122-125.
11. Antila H, Ryazantseva M, Popova D, et al. Isoflurane produces antidepressant effects and induces TrkB signaling in rodents. *Sci Rep* 2017;7.
12. Tremoleda JL, Sosabowski J. Imaging technologies and basic considerations for welfare of laboratory rodents. *Lab Anim* 2015;44:97-105.
13. Hoyer C, Gass N, Weber-Fahr W, et al. Advantages and challenges of small animal magnetic resonance imaging as a translational tool. *Neuropsychobiology* 2014;69(4):187-201.
14. Lauer AM, El-Sharkawy AM, Kraitchman DL, et al. MRI acoustic noise can harm experimental and companion animals. *J Magn Reson Imaging* 2012;36(3):743-747.
15. Houpt TA, Pittman DW, Barranco JM, et al. Behavioral effects of high-strength static magnetic fields on rats. *J Neurosci* 2003;23(4):1498-1505.
16. Ham CL, Engels JM, van de Weil GT, et al. Peripheral nerve stimulation during MRI: effects of high gradient amplitudes and switching rates. *JMRI-J Magn Reson Im* 1997;7(5):933-937.

17. Davids M, Guérin B, Vom Endt A, et al. Prediction of peripheral nerve stimulation thresholds of MRI gradient coils using coupled electromagnetic and neurodynamic simulations. *Magn Reson Med* 2019;81(1):686-701.
18. Schmale I, Gleich B, Rahmer J, et al. MPI safety in the view of MRI safety standards. *IEEE T Magn* 2015;51(2).
19. Glover PM. Interaction of MRI field gradients with the human body. *Phys Med Biol* 2009;54(21):R99-R115.
20. Medical Electrical Equipment—Part 2-33: Particular requirements for the basic safety and essential performance of MR equipment for medical diagnosis, IEC Standard 60601-2-33, 2003.
21. El Hamrani D, Vives V, Buchholz R, et al. Effect of Long-Term Retention of Gadolinium on Metabolism of Deep Cerebellar Nuclei After Repeated Injections of Gadodiamide in Rats. *Invest Radiol* 2020;55(2):120-128.
22. Wang S, Hesse B, Roman M, et al. Increased Retention of Gadolinium in the Inflamed Brain After Repeated Administration of Gadopentetate Dimeglumine: A Proof-of-Concept Study in Mice Combining ICP-MS and Micro- and Nano-SR-XRF. *Invest Radiol* 2019;54(10):617-626.
23. Aslakson CJ, Miller FR. Selective events in the metastatic process defined by analysis of the sequential dissemination of subpopulations of a mouse mammary tumor. *Cancer Res* 1992;52:1399-1405.

24. Wilhelm SM, Dumas J, Adnane L, et al. Regorafenib (BAY 73-4506): a new oral multikinase inhibitor of angiogenic, stromal and oncogenic receptor tyrosine kinases with potent preclinical antitumor activity. *Int J Cancer* 2011;129(1):245-255.
25. Kanzler S, Rix A, Czigany Z, et al. Recommendation for severity assessment following liver resection and liver transplantation in rats: Part I. *Lab Anim* 2016;50(6):459-467.
26. Touma C, Sachser N, Möstl E, et al. Effects of sex and time of day on metabolism and excretion of corticosterone in urine and feces of mice. *Gen Comp Endocrinol* 2003;130:267-278.
27. Rueden CT, Schindelin J, Hiner MC, DeZonia BE, Walter AE, Arena ET, Eliceiri KW. ImageJ2: ImageJ for the next generation of scientific image data. *BMC Bioinformatics* 2017;18(1):529.
28. European Parliament, Council of the European Union, 2010. Directive 2010/63/EU of the European Parliament and of the Council of 20 September 2010 on the protection of animals used for scientific purposes. Council of Europe, Strasbourg, 2010.
29. Davis HP, Mizumori SJ, Allen H, et al. Behavioral studies with mice exposed to DC and 60-Hz magnetic field. *Bioelectromagnetics* 1984;5(2):147-164.
30. Trzeciak HI, Grzesik J, Bortel M, et al. Behavioral effects of long-term exposure to magnetic fields in rats. *Bioelectromagnetics* 1993;14(4):287-297.
31. Murakami J, Torii Y, Masuda K. Fetal development of mice following intrauterine exposure to a static magnetic field of 6.3 T. *Magn Reson Imaging* 1992;10(3):443-437.

32. Ossenkopp KP, Innis NK, Prato FS, et al. Behavioral effects of exposure to nuclear magnetic resonance imaging: I. Open-field behavior and passive avoidance learning in rats. *Magn Reson Imaging* 1986;4:275-280.
33. Lockwood DR, Kwon B, Smith JC, et al. Behavioral effects of static high magnetic fields on unrestrained and restrained mice. *Physiol Behav* 2003;78(4-5):635-640.
34. You R, Liu Y, Chang RCC. A behavioral test battery for the repeated assessment of motor skills, mood and recognition in mice. *Jove-J Vis Exp* 2019;145.
35. Bronikowski AM, Carter PA, Swallow JG, et al. Open-field behavior of house mice selectively bred for high voluntary wheel-running. *Behav Genet* 2001;31(3):309-316.
36. Wilson DV, Evans AT, Carpenter RA, et al. The effect of four anesthetic protocols on splenic size in dogs. *Vet Anaesth Analg* 2004;31(2):102-108.
37. Jacobsen KO, Villa V, Miner VL, et al. Effects of anesthesia and vehicle injection on circulating blood elements in C3H/HeN male mice. *Contemp Top Lab Anim Sci* 2004;43(5):8-12.
38. DuPre SA, Hunter KW Jr. Murine mammary carcinoma 4T1 induces a leukemoid reaction with splenomegaly: association with tumor-derived growth factors. *Exp Mol Pathol* 2007;82(1):12-24.
39. Michel C, Neubauer A, Burchert A. Molekulare Tumorthherapie. *Der Internist* 2015;56(12):1389-1402.

40. Strelczyk D, Eichhorn ME, Luedemann S, et al. Static magnetic field impair angiogenesis and growth of solid tumors in vivo. *Cancer Biol Ther* 2009;8(18):1756-1762.

41. Kim R. Effect of surgery and anesthetic choice on immunosuppression and cancer recurrence. *J Transl Med* 2018;16(8).

42. Meier A, Gross ETW, Schilling JM, et al. Isoflurane impacts murine melanoma growth in a sex-specific, immune-dependent manner: a brief report. *Anesth Analg* 2018;126(6):1910-1913.

Tables

Table 1: Results of the rotarod test, heart rate and FCMs measurements in healthy BALB/c mice.

Values at day 27 of the experiment are given as % change to the baseline.

Parameter	Control	Isoflurane	1T MRI	7T MRI	CE 7T MRI
Rotarod [%]	126 ± 46	155 ± 39	142 ± 42	143 ± 47	97 ± 20
Heart rate [%]	108 ± 11	109 ± 22	124 ± 22	104 ± 21	103 ± 7
FCMs [%]	83 ± 24	98 ± 29	107 ± 29	60 ± 22	106 ± 37

Table 2: Results of the rotarod test, heart rate and FCMs measurements in 4T1 tumor-bearing BALB/c mice. Values at day 15 of the experiment are given as % change to the baseline.

Parameter	Vehicle			Regorafenib		
	Control	Isoflurane	CE MRI	Control	Isoflurane	CE MRI
Score [points]	5.7 ± 1.6	9.0 ± 5.5	8.2 ± 2.4	5.0 ± 1.6	9.9 ± 4.2	8.0 ± 2.5
Rotarod [%]	103 ± 30	114 ± 35	95 ± 35	118 ± 35	125 ± 30	98 ± 29
Heart rate [%]	106 ± 22	96 ± 34	100 ± 20	96 ± 26	98 ± 21	100 ± 15
FCMs [%]	99 ± 40	111 ± 44	110 ± 36	132 ± 88	132 ± 87	160 ± 26

Figures

Figure 1: Overview of A: experimental groups and B: timeline for healthy BALB/c mice to evaluate the influence of MRI on animal behavior and health state. FCMs = Fecal Corticosterone Metabolites. Animals were imaged thrice per week over a four weeks period with MRI sequences commonly used in drug response studies. Representative transversal images of the mammary fat pad acquired with C: a T_{1w} RARE and D: a T_{2w} RARE sequence. E: Representative MR image of a healthy mouse liver assessed with a T_{2w} RARE sequence.

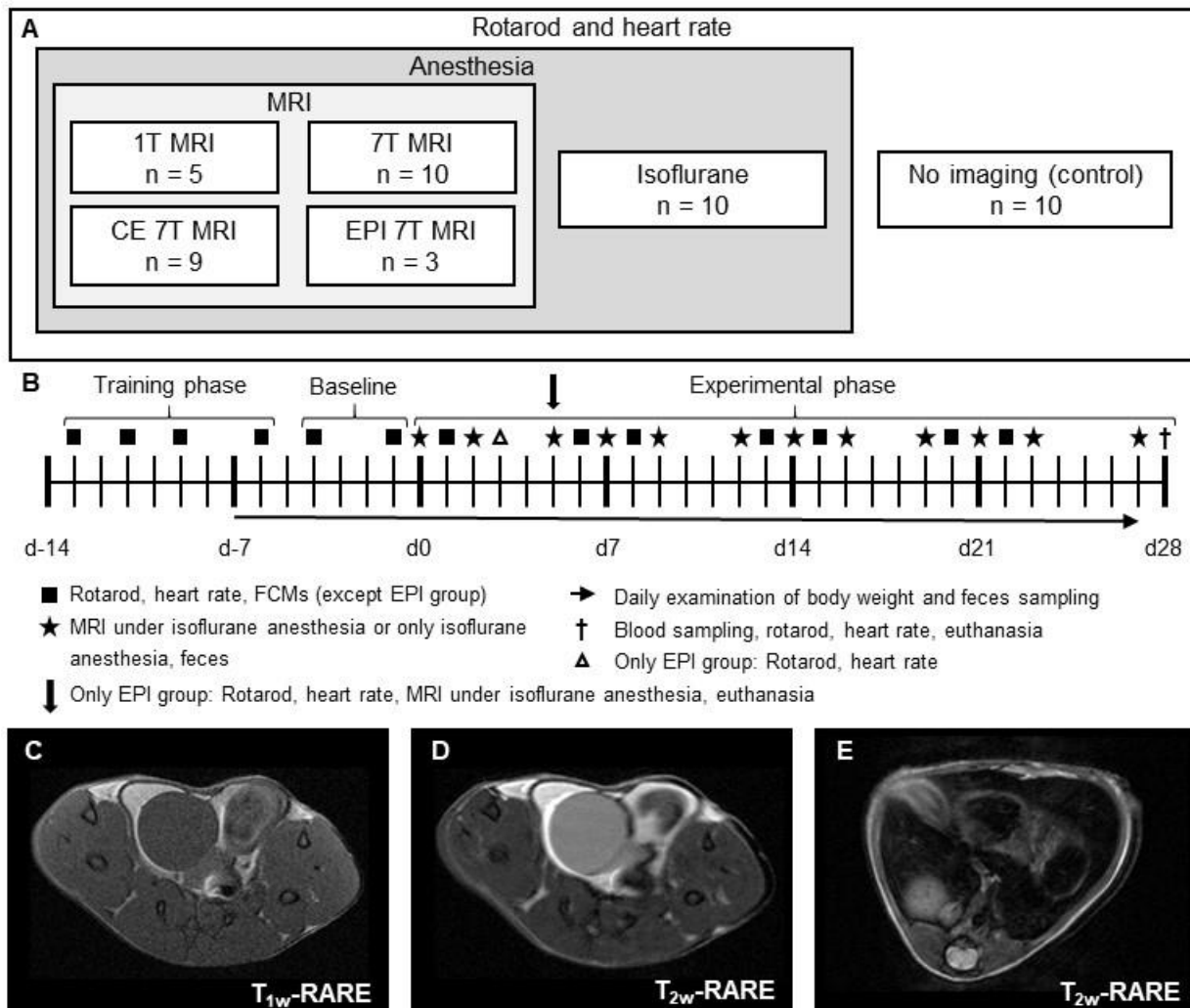


Figure 2: Overview of A: experimental groups and B: timeline for 4T1 tumor-bearing BALB/c mice. FCMs = Fecal Corticosterone Metabolites. The influence of isoflurane anesthesia and MR imaging with contrast agent (Gadovist, Bayer) on study results was investigated 3 times over two weeks in treated (regorafenib) and untreated (vehicle) 4T1 tumor-bearing mice with MRI sequences commonly used in drug response studies. Representative transversal images of a 4T1 tumor located in the mammary fat pad acquired using C: a T_{1w} RARE and D: a T_{2w} RARE sequence. Liver metastases were excluded by T_{2w} MR imaging using a RARE sequence. E: A representative MR image at the level of the liver.

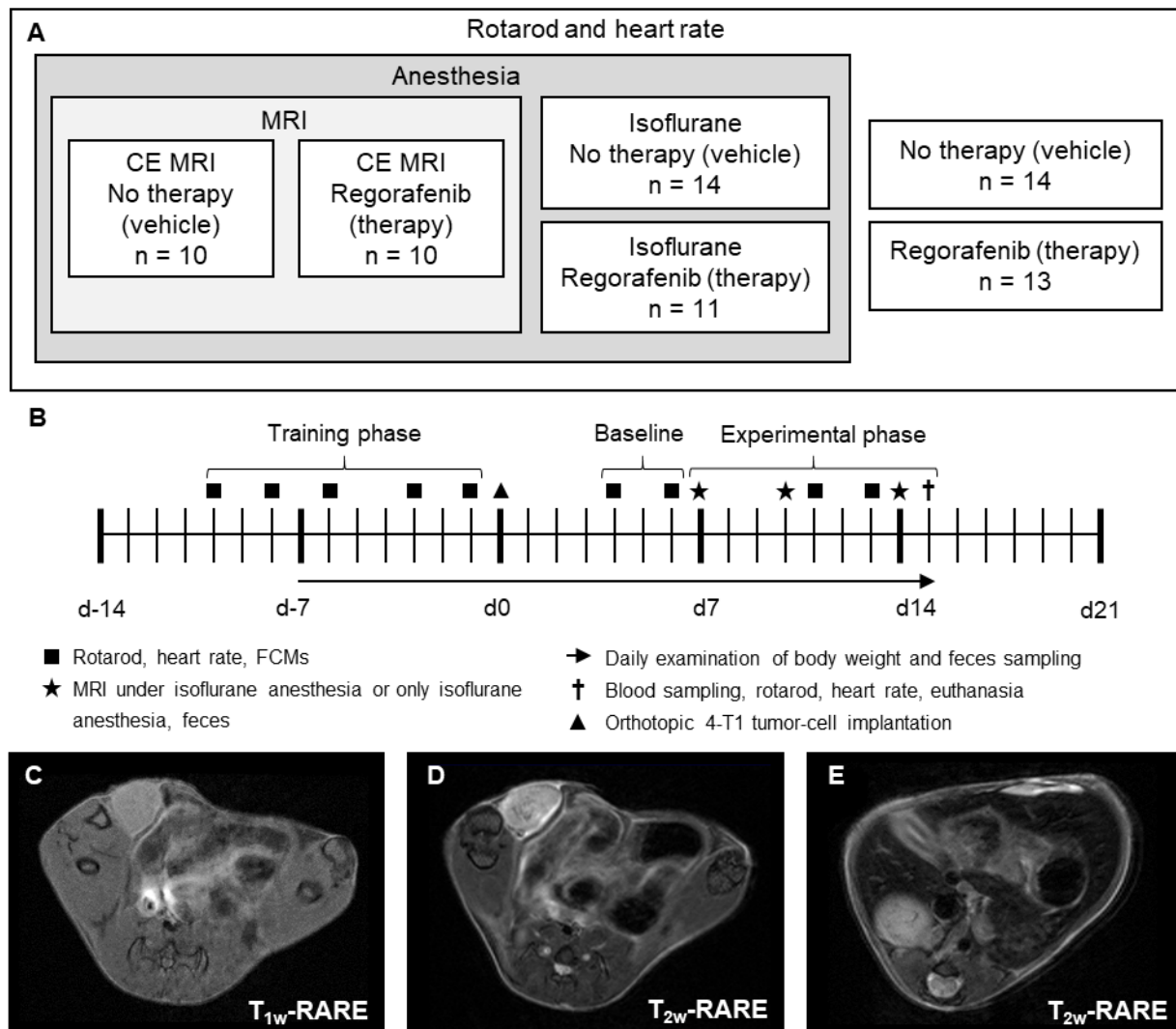


Figure 3: Influence of MRI on animal welfare, leukocyte counts and spleen weights (healthy BALB/c mice). BL = Baseline. A: The score sheet evaluation (based on body weight, fur appearance, body openings, behavior, body temperature and heart rate) shows a mild burden for all animals independent of MRI. B: Longitudinal rotarod evaluation (rotations per minute (rpm)) suggests low stress levels in mice of all groups independent of the performance of MRI scans. C: Longitudinal heart rate measurements indicate no change over time. D: Fecal corticosterone metabolites (FCMs) (stress hormone levels) are stable in all groups over time. E: Repeated MRI examinations have no influence on leukocyte counts. F: Spleen weights are reduced in all anesthetized groups ($p < 0.001$), but not further affected by MRI examinations.

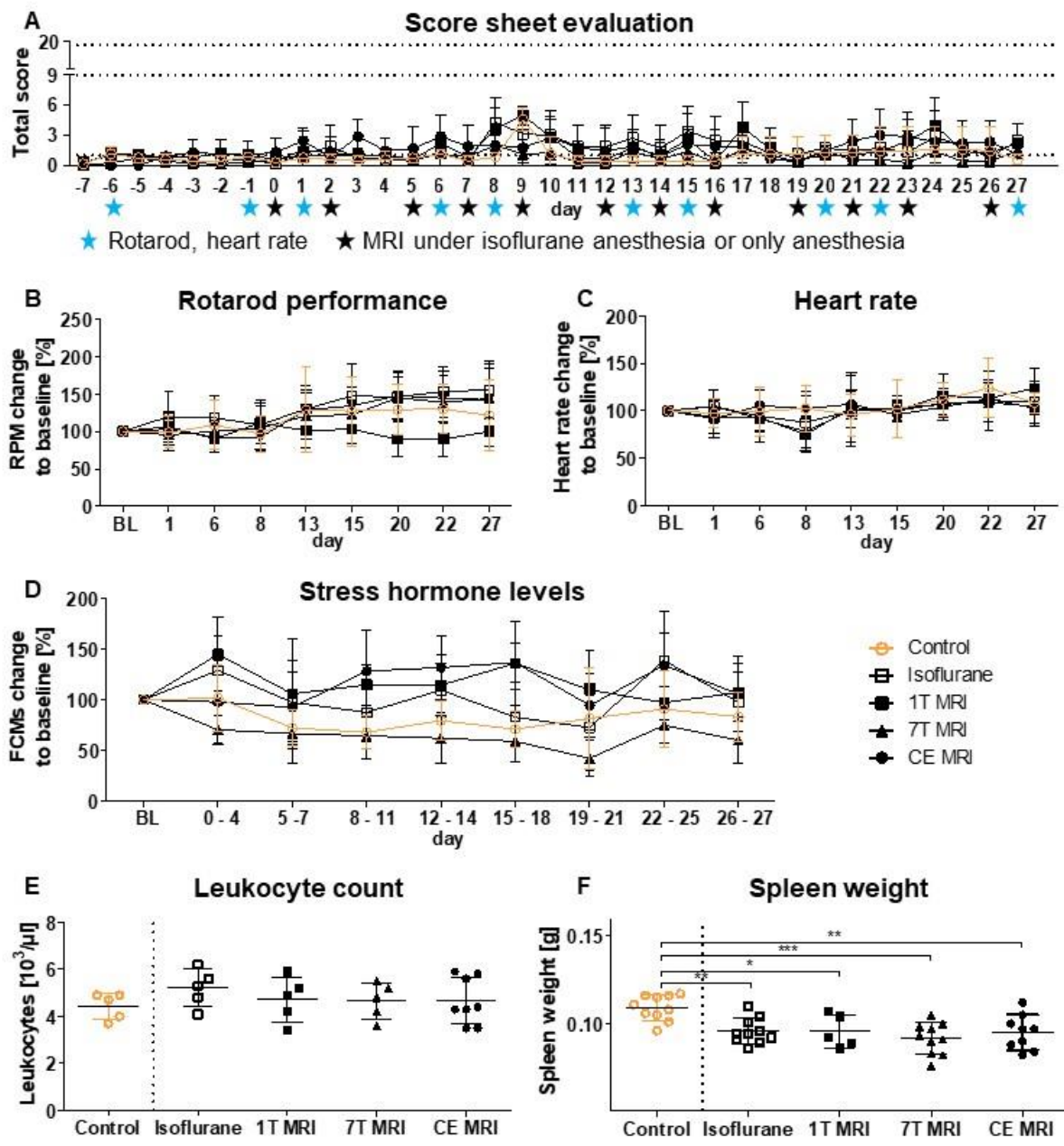


Figure 4: Influence of MRI on therapy response of 4T1 tumors in BALB/c mice. Isofl. = Isoflurane.

A: T_{2w} MRI image (7T) of a 4T1 tumor located in the mammary fat pat. B: CE MR imaging and anesthesia have no influence on tumor growth in untreated and treated animals. Dotted lines indicate vehicle and regorafenib control groups. C: Staining of perfused vessels with CD31 and lectin does not indicate differences between the groups. D: Macrophage (F4/80 staining) infiltration in tumors does not change if isoflurane or CE MRI scans are performed. E: TUNEL staining indicates no differences in apoptotic cell rates in tumors of vehicle and regorafenib treated mice after isoflurane anesthesia or CE MRI. Scale bar: 100 μ m (images taken at 20x magnification).

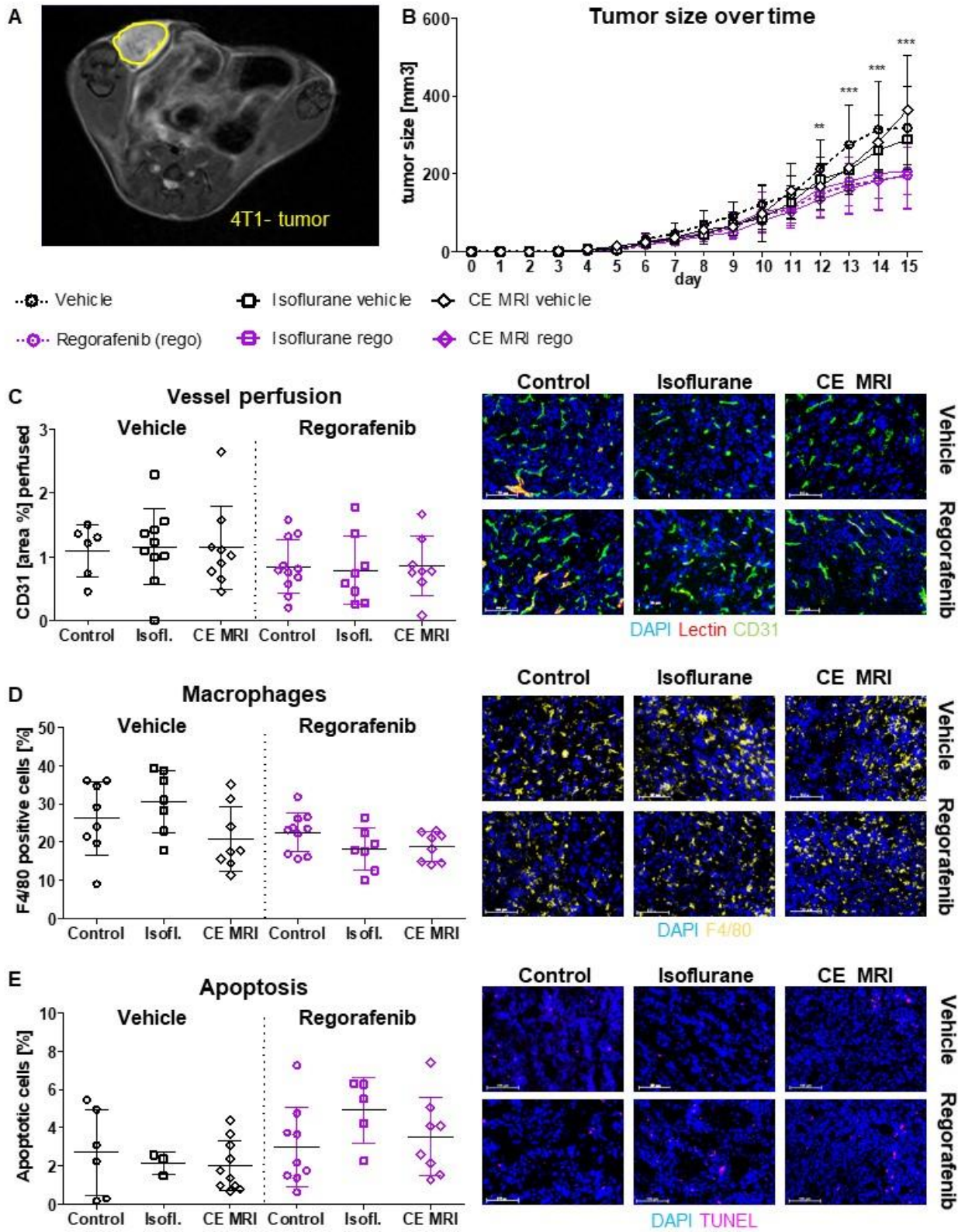
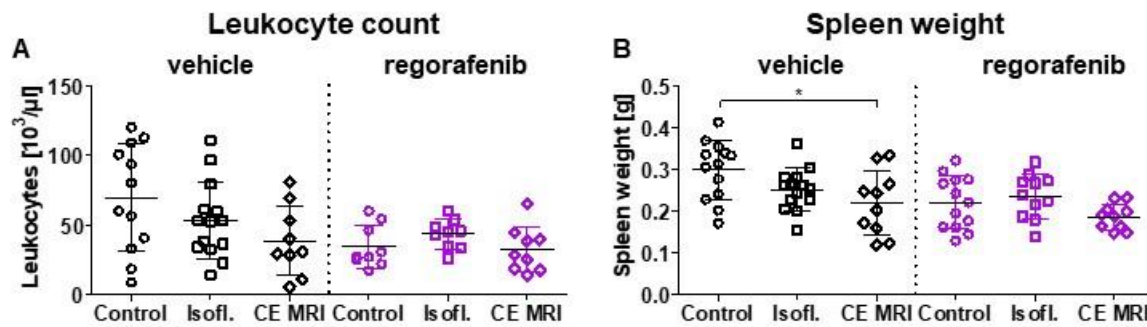


Figure 5: Influence of CE MRI on leukocyte counts and spleen weights in 4T1 tumor-bearing BALB/c mice. Isofl. = Isoflurane anesthesia A: The highest leukocyte counts are found in untreated control animals. Isoflurane and CE MRI exposure tend to decrease leucocyte levels in untreated animals. Regorafenib treated animals have lower leukocyte concentrations at the end of the experiment. However, there is no difference between the regorafenib treated control group and the regorafenib treated isoflurane and CE MRI exposed groups. B: In untreated animals, spleen weights are reduced after isoflurane anesthesia and significantly lower in animals after CE MRI examinations ($p=0.022$). In regorafenib treated animals spleen weights are lower than in untreated control animals. However, no significant differences between the control, isoflurane and CE MRI groups are found.



Influence of MRI examinations on animal welfare and study results

Supplemental Digital Content

Material and Methods

Magnetic resonance imaging protocol

Table S1: 1T MRI standard scan protocols. TR = Repetition time, TE = Echo time, NSA = Number of signal averages, FOV = Field of view. Imaging was performed using a RF RES 42.5 1H 030/80 LIN TR coil.

Sequence/ Parameter	T1 RARE	T2 RARE	T1 FLASH
TR	585 ms	1699 ms	111 ms
TE	12 ms	84 ms	6 ms
Echo spacing	12 ms	28 ms	
Rare factor	2	8	
Flip angle	90°	90°	30°
NSA	8	8	1
FOV	35 x 35 mm	35 x 35 mm	30 x 30 mm
Matrix	256 x 256	192 x 192	54 x 64
Slice thickness	1.25 mm	1.25 mm	1.25 mm
Pixel size	137 x 137 µm	182 x 182 µm	469 x 469 µm
Scan time	7:29 min	5:26 min	10:04 min for 80 measurements

Table S2: 7T MRI standard and EPI scan protocols. TR = Repetition time, TE = Echo time, NSA = Number of signal averages, FOV = Field of view. Imaging was performed using a RF RES 300 1H 075/040 QSN TR coil.

Sequence/ Parameter	T1 RARE	T2 RARE	T1 FLASH	EPI 1	EPI 2	EPI 3
TR	1500 ms	2500 ms	2015 ms	667 ms	300 ms	900 ms
TE	8 ms	40 ms	175 ms	31 ms	22 ms	39 ms
Echo spacing	8 ms	13 ms				
Rare factor	4	8				
Flip angle	90°	90°	30°	90°	90°	90°
NSA	2	8	1	44	10	30
FOV	32 x 22 mm	32 x 25 mm	32 x 25 mm	20 x 20 mm	12 x 8 mm	20 x 20 mm
Matrix	256 x 256	200 x 200	64 x 64	96 x 80	64 x 50	160 x 160
Slice thickness	1.0 mm	1.0 mm	1.0 mm	0.7 mm	0.8 mm	0.5 mm
Pixel size	123 x 86 μ m	158 x 123 μ m	500 x 391 μ m	208 x 250 μ m	188 x 160 μ m	125 x 125 μ m
Scan time	2:24 min	8:20 min	11:12 min for 80 measurements	1:00 min	1:00 min	1:00 min

Table S3: Score sheet for healthy and tumor-bearing mice. Alterations in body weight, general state, spontaneous behavior, and treatment specific parameters were documented and allocated to a point grading system, where 0 points describe no alteration of the physiological state and ≥ 20 points the highest severity and humane endpoint. For tumor-bearing mice tumor growth and body conditioning score were additionally scored.

Parameters for healthy and tumor-bearing mice	
Observation	Score
I Body weight	
No changes	0
Change < 5%	1
Weight reduction 5 – 10 %	5
Weight reduction 11 – 19 %	10
Weight reduction ≥ 20 %	20
III General condition	
Shiny fur, clean body openings, clear and shiny eyes	0
Defects of the fur, (increased or decreased body care)	1
Dull fur, untidy, unclean body openings, cloudy eyes, increased muscle tonus	5
Dirty fur, clotted and soggy body openings, unusual posture, dull eyes, high muscle tonus	10
Cramps, paralysis (trunk muscles, extremities), conspicuous breathing problems, cold body	20
IV Behavior	
Normal behavior (sleep, reaction to blowing and touching, curiosity, social contacts)	0
Slight changes to normal behavior	1
Unusual behavior, impaired motor function or hyper kinetics	5
Self-isolation, lethargy, pronounced hyper kinetics or behavioral stereotypes, coordination disorders	10
Pain sounds when seizing, self-amputation (autoaggression, autotomy), apathy, noticeable defensive reactions	20
V Clinical symptoms	
Temperature and heart rate normal, mucosa well supplied with blood	0
Slight change to the normal situation	1
Change of body temperature from 1-2 °C, pulse + 30	5
Change of the body temperature from > 2 °C, pulse + 50	20
Additional parameters for tumor-bearing mice	
VI Body Conditioning Score	
Spine and pelvis not prominent (BC3)	0
Spine prominent, segmentation of vertebral column evident, pelvis palpable (BC2)	10
Skeletal structure and vertebral column highly prominent, little or no subcutaneous fat (BC1)	20
VII Orthotopic tumor growth	
Tumor $\varnothing > 5$ mm to < 10 mm	5
Tumor $\varnothing > 10$ mm to < 15 mm	10
Tumor $\varnothing > 15$ mm or tumor size > 10% of body weight	20

Neuronal effects (motion uncertainty, loss of sensitivity (no reaction to touch))	10
Exulceration, invasive tumor growth, ascites or bleeding	20
Highly enlarged spleen or lymph nodes (imaging)	20
Soft stool	5
Persistent diarrhea	20
Assessment	Total score
Degree of stress 0: no stress	0
Degree of stress 1: low stress, continue to observe carefully	1 – 9
Degree of stress 2: moderate stress, if necessary initiate veterinary case (analgesia)	10 – 19
Degree of stress 3: high stress, remove animal form the study	20
Humane end point	≥ 15 for more than 48 hours

Table S4: Hemograms of healthy mice (\pm standard deviation). Comparison of all groups to control: $p > 0.05$.

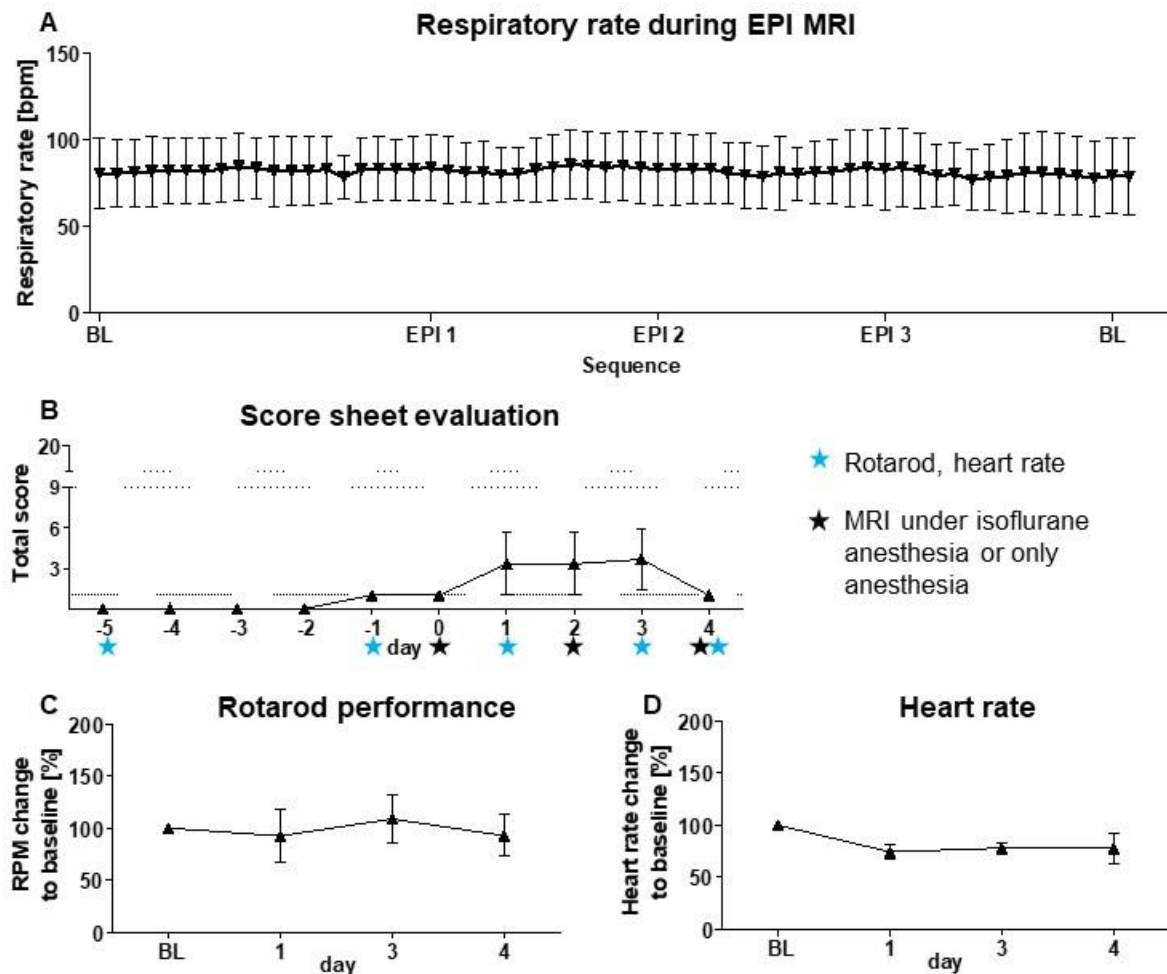
Healthy mice	Control	Isoflurane	7T MRI	1T MRI	CE 7T MRI
Leukocytes [$10^3/\mu\text{l}$]	4.44 \pm 0.56	5.20 \pm 0.80	4.66 \pm 0.77	4.72 \pm 0.96	4.66 \pm 0.98
Erythrocytes [$10^6/\mu\text{l}$]	8.85 \pm 0.14	9.21 \pm 0.38	9.30 \pm 0.24	9.46 \pm 0.27	8.86 \pm 0.48
Hemoglobin [g/dL]	14.00 \pm 0.48	14.46 \pm 0.28	14.72 \pm 0.49	14.90 \pm 0.46	14.66 \pm 1.06
Hematocrit [%]	37.44 \pm 0.86	38.56 \pm 1.35	38.94 \pm 0.91	39.48 \pm 0.52	39.69 \pm 2.26
Thrombocytes [$10^5/l$]	10.38 \pm 0.80	10.53 \pm 0.65	11.47 \pm 0.48	10.53 \pm 0.64	9.00 \pm 0.11

Table S5: Hemograms of 4T1 tumor bearing mice (\pm standard deviation). Comparison between vehicle groups to control: $p>0.05$; comparison between regorafenib groups to control: $p>0.05$.

4T1-bearing mice	Vehicle			Regorafenib		
	Control	Isoflurane	CE MRI	Control	Isoflurane	CE MRI
Leukocytes [$10^3/\mu\text{l}$]	69.45 \pm 38.58	53.07 \pm 27.38	38.57 \pm 25.09	34.33 \pm 15.26	43.34 \pm 10.48	32.32 \pm 16.40
Erythrocytes [$10^6/\mu\text{l}$]	8.81 \pm 0.79	8.96 \pm 0.53	8.89 \pm 0.37	9.02 \pm 0.67	9.01 \pm 0.77	8.99 \pm 0.38
Hemoglobin [g/dL]	14.79 \pm 1.37	14.83 \pm 1.03	14.99 \pm 0.60	13.93 \pm 2.02	15.01 \pm 1.70	14.87 \pm 0.88
Hematocrit [%]	39.72 \pm 3.87	39.81 \pm 2.56	40.26 \pm 1.41	37.30 \pm 5.52	39.40 \pm 3.68	39.60 \pm 1.75
Thrombocytes [$10^5/\text{l}$]	6.19 \pm 0.70	6.21 \pm 0.12	6.81 \pm 0.15	6.22 \pm .016	6.10 \pm 0.90	6.30 \pm 0.77

Influence of EPI MRI on animal welfare and respiratory rate

Figure S1: Investigation of acute and long-term Echo-Planar-Imaging (EPI) effects (for sequence information see supplemental material) on animal welfare and respiratory rate in BALB/c mice. BL = Baseline. A: The respiratory rate, documented every 5 second, is stable during EPI measurements. Bpm = Breaths per minute. B: The score sheet evaluation (body weight, fur appearance, body openings, behavior, body temperature and heart rate) shows only a mild burden for the animals after EPI MRI. C: Rotarod performance and D: Heart rates are not influenced by EPI MRI.



Influence of CE MRI on animal welfare in 4T1 tumor-bearing mice

Figure S2: Influence of 7T CE MRI on animal welfare in 4T1 tumor-bearing BALB/c mice. BL = Baseline. Dotted lines indicate the vehicle and regorafenib control groups. A: The score sheet evaluation (based on body weight, fur appearance, body openings, behavior, body temperature and heart rate, tumor growth and body conditioning score) shows a mild, in some individuals moderate, burden for the animals that is not influenced by contrast enhanced MRI in untreated and treated groups and based on tumor progression. B: Rotarod performances are not altered after repeated CE MRI. C: Imaging has no long-term effects on animals' heart rate. D: Fecal corticosterone metabolites (FCMs) are not increased in mice, even after repeated imaging.

